

Transdermal Histamine in Multiple Sclerosis, Part Two: A Proposed Theoretical Basis for Its Use.

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Abstract

This paper is the companion to an earlier publication, which discussed preliminary results of transdermal histamine use for ameliorating symptoms of both relapsing-remitting and progressive multiple sclerosis (MS). Here we include preliminary findings on the impairments of digestion and assimilation in MS patients seen in a private clinic. Although only a small number of patients was surveyed, an association was found between impaired gastric acid production, impaired protein hydrolysis, and subnormal plasma histidine levels in patients with MS. Impaired digestion might, therefore, impair the ability of MS patients to synthesize histamine. This paper discusses how impairment of histamine synthesis might lead to symptoms of MS, and conversely how exogenously administered histamine might alleviate symptoms. Various mechanisms of action are suggested, including: enhanced gastric acid and pancreatic enzyme secretion, augmentation of subnormal cerebral tissue levels of histamine, improved electrical function of demyelinated fibers, increased cerebral blood flow, suppression of aberrant autoimmune responses, and stimulation of remyelination. We also discuss the observed failure of digestive function in MS and point out that pathological changes which parallel CNS findings have been found in the enteric nervous system (ENS) of patients with Parkinson's disease. Similar parallels might exist between the CNS and ENS in multiple sclerosis.

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Introduction

A previous paper¹ discussed preliminary results of usage of a transdermal histamine/caffeine preparation for ameliorating symptoms of multiple sclerosis (MS). Improvement was seen in 37 of 55 patients (67%) followed for at least six weeks. This work has continued, and at the time of writing 167 patients had been followed for at least six weeks, with 26 percent of patients experiencing significant improvement as defined previously,¹ 29 percent experiencing some improvement, and 45 percent seeing no benefit. Improvements at three months have, in most cases, continued at six months.

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Areas of improvement have been observed in extremity strength, balance, bladder control, fatigue, heat tolerance, cognitive function, peripheral edema, and activities of daily living. Improvement is often seen within a few hours or days of starting therapy, and there have been minimal side-effects. This is in keeping with the findings of others who have administered similar amounts of subcutaneous histamine for such clinical problems as vertigo, headache,² and cancer.³

Five possible mechanisms of action of histamine on the symptoms and clinical manifestations of MS were introduced in Part One of this article.¹ These include augmentation of subnormal cerebral tissue levels of histamine, improved electrical function of demyelinated fibers, increased cerebral blood flow, modulation of autoimmune responses, and stimulation of remyelination.

Although the respective bodies of literature on histamine and MS are vast, there are no recent studies exploring histamine's role in the treatment of MS, and only a few studies even tangentially examine the role of histamine in the pathophysiology of MS. A recent Medline search under the terms "histamine and multiple sclerosis" returned 16 citations, with only some of these papers relevant to the issues discussed here. Consequently, we emphasize that the ideas discussed here are mostly speculative and unsupported by direct experimentation. Should histamine therapy continue to show promise, we hope other researchers will use this paper as a starting point for testing these hypotheses.

Histamine Neurophysiology and Neuroanatomy

There is considerable evidence in the literature that histamine is an ancient and pervasive regulator of physiologic processes. Histaminergic neural paths extend throughout the bodies of lower organisms, including insects⁴ and marine animals.⁵ Histaminergic fibers are

found in the sympathetic chains of mice⁶ and higher mammals.

Histaminergic neural paths extend from the brain to synapses in the higher cervical spinal cord.⁷ In humans, histaminergic fibers project from a dense population of cell bodies in the hypothalamus to almost all areas of the brain, including the cerebellum.⁸⁻¹⁰ The goat pineal gland contains a large amount of histamine.¹¹ This suggests a link between histamine and melatonin production, although similar data on humans has not been obtained. Histamine is synthesized within these neurons by decarboxylation of the amino acid histidine.

The hypothalamic histaminergic system modulates diverse and crucial aspects of physiology, including thirst, hunger, sex drive, circadian rhythm, arousal level,¹² urine output,^{13,14} thermoregulation,¹⁵ nociception,¹² and vestibular function.^{16,17} Histamine also participates in other aspects of neuroendocrine function, including stress-induced secretion of such mediators as adrenocorticotropin (ACTH), beta endorphin, melatonin, prolactin, and peripheral catecholamines.^{18,19}

Three different receptor subtypes, denoted H1, H2, and H3, mediate these responses. Discussion of these receptors can be found in reference 20. Briefly, all the histamine receptors are thought to be G-protein coupled receptors. Stimulation of the H1 receptor is, in general, associated with an increase in intracellular Ca^{2+} and formation of inositol triphosphate and diacylglycerol. Stimulation of the H2 receptor results in an elevation of intracellular cyclic adenosine monophosphate (cAMP). The intracellular coupling of the H3 receptor is not well understood; possible mechanisms include coupling to calcium channels. On neurons, the H3 receptor is a presynaptic receptor that is inhibitory both for release of histamine (autoreceptor) and other neurotransmitters (heteroreceptor).

The interpretation of experimental findings pertaining to histamine is sometimes

difficult due to the number of histamine receptors and their diverse actions. For a given phenomenon, the action of histamine at H1 receptors often opposes that seen at H2 receptors.²¹ Sometimes the effects of stimulation at both receptors are the same.¹⁸ Furthermore, H3 receptor stimulation inhibits the release of histamine from mast cells, including brain mast cells.²² The effect of histamine in any given situation may, therefore, be sensitive to the relative populations of the three receptor subtypes at a given location, as well as the concentration of histamine in the vicinity of the receptors.

The following examples illustrate the diversity of action of histamine in the brain. In rats, H1 stimulation decreases feeding,¹³ increases alertness, and increases nociception,¹² whereas H3 receptor stimulation provokes drinking.¹³ Both H1 and H2 stimulation (via intracerebroventricular histamine receptor agonist injection) elicited hypothermia in mice, whereas H3 stimulation prevented development of hypothermia.¹⁵ Intracerebroventricular injection of histamine also stimulates diuresis in rats¹³ via H2 receptors. Histamine also stimulates release of melanocyte stimulating hormone from the pituitary via both H1 and H2 receptor stimulation.¹⁸

Approximately one-half of the histamine in the mouse brain resides in mast cells, as shown by Maeyama, who studied mutant mice devoid of mast cells.²³ Outside the brain, considerable histamine is stored in the enterochromaffin-like (ECL) cells of the gastric mucosa, where histamine plays a critical role in acid secretion. Histamine is also found in circulating basophils, eosinophils and platelets, as well as mast cells in many tissues, including the skin, heart, lungs, liver, kidneys, spleen, bladder, ovaries, and testicles.²⁴⁻²⁷ Further discussion of the role of histamine in these tissues is beyond the scope of this article.

Pathophysiology of MS

The majority of current research supports the view that MS is an autoimmune disorder in which immune cells (T lymphocytes and macrophages) in the blood are “primed” (sensitized), possibly by viral-related antigens, to attack myelinated neurons and glial cells of the central nervous system.^{28,29} Candidate viruses include measles,³⁰ herpes,³¹ vaccinia,³² and multiple sclerosis retrovirus.^{33,34}

The concept of Th1/Th2 balance is central to any discussion of the immunologic aspects of MS. Briefly, many diseases recognized as autoimmune can be classified according to the activity of two T cell subpopulations: Th1 and Th2.³⁵ Differentiation of naive CD4+ T cells to a Th1 phenotype is induced by interleukin-12 (IL-12).³⁶ Th1-polarized cells up-regulate various aspects of cell-mediated immunity by the secretion of cytokine mediators, including IL-1, α IL-1, β IL-2, IL-8, IL-12, tumor necrosis factor alpha (TNF α), and interferon gamma (IFN γ). Th1-dominant conditions include infection by intracellular pathogens, rheumatoid arthritis (RA), Crohn’s disease, autoimmune thyroiditis, and delayed hypersensitivity reactions.

Differentiation of naive T cells to a Th2 phenotype is primarily promoted by IL-4.³⁶ Th2-polarized cells secrete mediators including IL-4, IL-5, IL-10, and IL-13, which up-regulate humoral immune responses commonly regarded as “allergic,” including IgE production and eosinophil function.³⁶ Th2-dominated conditions include parasitic infections, atopic dermatitis, asthma, systemic lupus erythematosus, and progressive systemic sclerosis. Pregnancy is also considered to be a Th2 dominated state.³⁵ Th2 cells may also control the development and activity of Th1 cells.³⁷

Immunochemical factors, including IL-1, granulocyte-macrophage colony stimulating factor (GM-CSF), TNF α and others act to bias naive CD4+ cells toward a Th1 state by stimulating production of IL-12 from antigen-presenting cells, including monocytes and

dendritic cells. Other factors, including IL-10, histamine, corticosteroids, vitamin D3, and beta agonists inhibit production of IL-12 from these same cells. Nagelkerken has suggested that hypothalamic-pituitary-adrenal (HPA) axis dysfunction, with deficient corticosteroid production, may lead to Th1 dominant states.³⁷

MS is regarded as a Th1 dominant state, and Beck³⁸ reported increased Th1 cytokine (IL-1, TNF α) secretion prior to relapse. Van Boxel-Dezair³⁹ demonstrated increased IL-12 mRNA and decreased IL-10 mRNA in MS patients compared to controls, and IL-12 mRNA levels correlated with disease activity. Administration of Copolymer 1 (Copaxone) has been shown to rebalance the immune response in MS by encouraging Th2 responses.^{40,41} In contrast, administration of interferon beta (IFN β) actually promotes IFN α production (Th1 response), yet IFN α is beneficial for some MS patients.⁴² Also, evidence of increased activity of Th2 cells in MS is seen in the form of elevated levels of autoantibodies.⁴³ This illustrates that although the Th1/Th2 paradigm is very useful, it is not a complete description of immune dysregulation in MS.

Accepting that “priming” of the immune system occurs, permeability of the blood-brain barrier is an important factor to consider, since no nerve damage occurs unless the primed cells gain access to the central nervous system. Spatial and temporal correlations between breaches in the blood-brain barrier and subsequent development of MS lesions support the central role of changes in permeability of the blood-brain barrier.^{44,45} Kwon demonstrated that longstanding plaques exhibit evidence of permanent damage to the blood-brain barrier.⁴⁶

Various factors known to alter the permeability of the blood-brain barrier have been directly or epidemiologically linked to the initiation of MS, or are associated with relapse. Examples include thiamine deficiency,⁴⁷ heavy metal toxicity,⁴⁸ and heat stress.⁴⁹ A good summary of some of the evidence supporting the

importance of blood-brain barrier permeability in MS is given by Wallace⁵⁰ along with a discussion of the evidence supporting the use of various supplemental nutrients to improve the integrity of the blood-brain barrier.

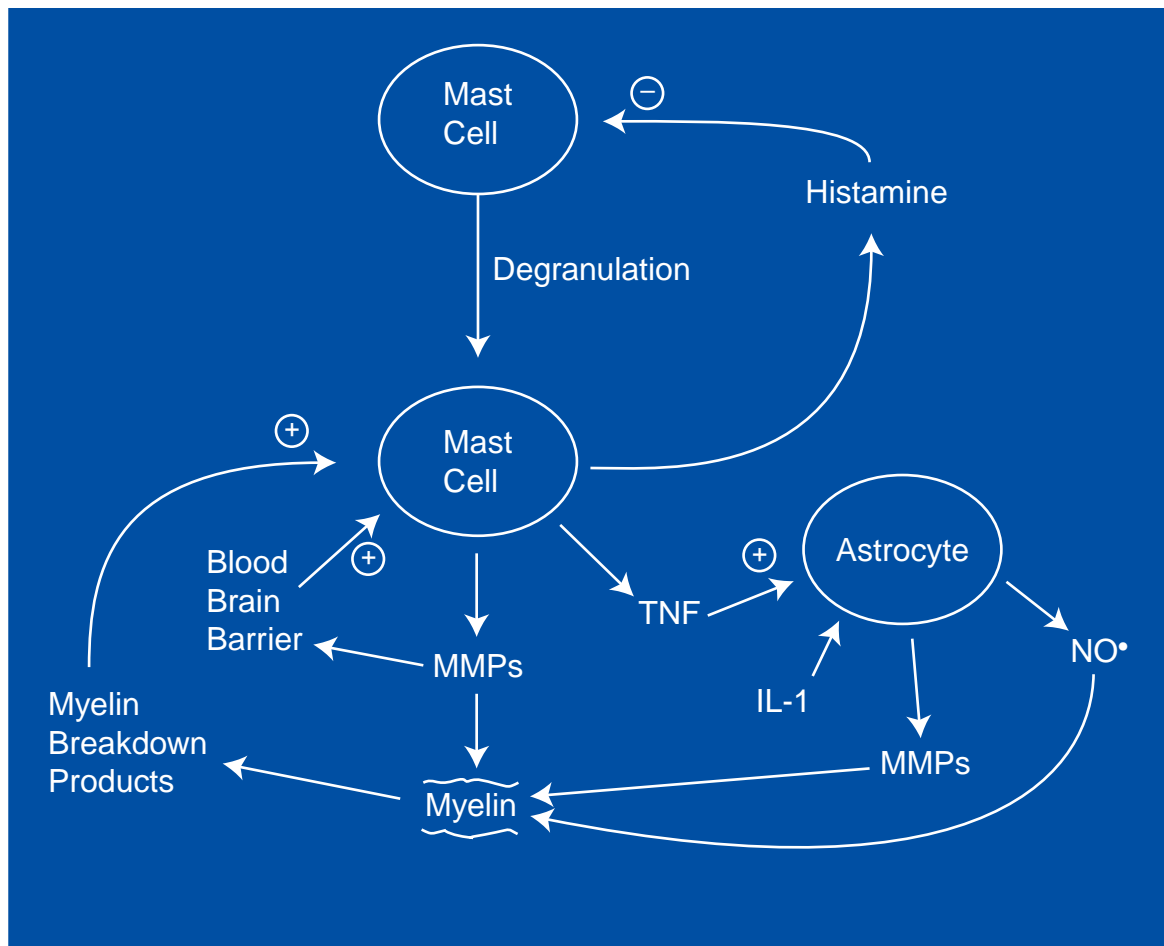
The microscopic appearance of MS plaques changes with time, but in both acute and chronic MS there is an inflammatory reaction dominated by T lymphocytes and macrophages. Myelinated fibers are engulfed by macrophages⁵¹ while other cells, such as peripheral lymphocytes and plasma cells, are also seen in close contact with myelin.⁵² In fresh lesions, demyelination is accompanied by destruction of other tissue elements, including oligodendrocytes, astrocytes, and axons. This destruction ends within a few weeks in many cases, and remyelination may follow, once the lesion has been repopulated with oligodendrocytes.⁵³

Blakemore⁵⁴ and Franklin⁵⁵ conclude that remyelination depends on migration of oligodendrocyte progenitors, such as those observed by Prineas,⁵⁶ from nearby undamaged tissue into the area of inflammation. Franklin also suggests extensive, recurrent, or longstanding inflammation will deplete surrounding healthy tissue of oligodendrocyte progenitors, ultimately limiting the repopulation of plaques and the remyelination of damaged axons.

Stimuli to remyelination include growth factors, such as platelet-derived growth factor,⁵⁷ triiodothyronine (T3),^{58,59} and vitamin B12.^{60,61} Elevation of intracellular cAMP has also been shown to play a role in induction of oligodendrocyte progenitor differentiation and myelin synthesis in the rat brain⁶² and in peripheral Schwann cells.^{63,64}

Thiamine also plays a role in myelination. Demyelination is a hallmark of thiamine deficiency,⁶⁵ although deficiencies of other nutrients such as copper⁶⁶ also cause demyelination. Thiamine, in the form of thiamine pyrophosphate, helps stabilize nerve cell membranes.⁶⁷ Myelin-synthesizing oligodendrocyte

Figure 1. Role of Mast Cell Mediators in CNS Inflammation.



cell bodies contain thiamine pyrophosphatase (TPPase),⁶⁸ the enzyme which generates thiamine pyrophosphate. TPPase has been localized to the Golgi complexes of various neuronal cells,⁶⁹ and Trapp found protein zero (P0), the main component of myelin basic protein, in Golgi complex membranes in rat Schwann cells.⁷⁰

The inflammatory cascade leading to demyelination is complex, and current understanding of it is by no means complete. There has been increased awareness lately of the important role of mast cells in central nervous system (CNS) inflammatory processes, including MS.^{71,72} Substances released from mast cells can attack myelin, and myelin breakdown products can stimulate further mast cell degranulation.⁷³ Skaper has suggested that down-regulation of mast cell activation could be a

therapeutic strategy in neuroinflammatory conditions,⁷⁴ and proxicromil, a mast cell stabilizer, has been employed successfully in experimental autoimmune encephalomyelitis (EAE), the mouse model of MS.⁷⁵

Mast cells are known to release leukotrienes,⁷⁶ inflammatory mediators that affect vascular permeability and exhibit chemoattractant properties. Elevated levels of leukotriene C4 and leukotriene B4 are found in the cerebrospinal fluid (CSF) of MS patients.⁷⁷ Inhibition of 5-lipoxygenase, an enzyme involved in the synthesis of leukotrienes, prevented the development of symptoms in guinea pig EAE and significantly reduced histologic inflammation scores.⁷⁸

Matrix metalloproteinases (MMPs), a family of zinc-containing enzymes which can digest connective tissue and myelin,^{72,79} are

another important class of inflammatory mediators. They are also known to be disruptive to the blood-brain barrier.⁸⁰ Mast cells contain and release MMPs,^{72,79,81} and MMPs are found to be elevated both in the CSF of MS patients⁸² and in EAE mice.⁸³ Blockage of MMPs inhibits or reduces the severity of EAE.^{83,84}

Earlier, reference was made to “priming” of peripheral immune cells. An example of this is the observation that peripheral monocytes and macrophages contain elevated levels of MMPs.^{80,82,83} The release of MMPs by mast cells may feed the inflammatory spiral in several ways. First, by generating myelin fragments such as myelin basic protein, which elicit further mast cell degranulation,^{74,79,85} and second, by liberating cytokines such as TNF α from the cell membranes of monocytes⁸⁶ or myelin autoreactive CD4+ T cells.⁸⁴ TNF α levels in the CSF of MS patients have been correlated to disease severity.^{87,88} Blockade of TNF by anti-TNF antibodies prevented the development of EAE in one mouse study.⁸⁹

Mast cells also release TNF α directly^{71,74} and mast cell-derived TNF affects the release of neurotoxic NO radicals (from astrocytes in mast cell/hippocampal co-cultures)⁹⁰ and MMPs.⁹¹ Thus, as mentioned, mast cell degranulation, with release of mediators such as MMPs and TNF, might set up a potential positive feedback loop (Figure 1) promoting more mast cell degranulation.

Histamine is the most widely recognized mediator released by mast cells. Release of histamine by brain mast cells may initially accelerate inflammation by increasing the permeability of the blood-brain barrier, an H2 receptor-mediated phenomenon.⁹² Increased blood-brain barrier permeability would then presumably increase the influx of sensitized peripheral immune cells. This process is thought to be central to the pathophysiology of MS. Ongoing mast cell degranulation with histamine release might also create an increased demand for histidine (the direct precursor of histamine), and

potentially compromise the supply of histidine at other histamine-synthesizing sites in the brain, such as histaminergic neurons.

Interestingly, the number, location, and histamine content of mast cells in mouse brain have been shown to be inherited traits.^{93,94} If this is also true in humans, some individuals might exhibit increased susceptibility to MS and other inflammatory conditions because of this genetic programming.

Gut Function in MS

Constipation is a common complaint of MS sufferers. For example, 68 percent of 280 patients surveyed by Weber⁹⁵ reported constipation or fecal incontinence. These symptoms may reflect CNS damage as well as enteric nervous system (ENS) dysfunction.⁹⁶ (Reference 96 is an overview of the ENS.)

Other aspects of gut function impairment in MS are less well recognized. These include impaired digestion and assimilation of nutrients. Gupta⁹⁷ reported evidence of fat malabsorption and protein maldigestion in 40 percent of 52 MS patients. Xylose malabsorption was seen in 27 percent, and B12 malabsorption in 12 percent. Absorption of fat-soluble vitamins A and beta carotene was also impaired to a certain extent. Iarosh noted endoscopic atrophic changes, gastritis, and ulcerations consistent with achlorhydria in 32 MS patients.⁹⁸ The total number of MS patients in that study was 89, but selection criteria for endoscopy were not given.

Over the past 25 years, one of the authors (JW) has observed abnormal gastric acid secretion, as assayed by Heidelberg capsule,⁹⁹ in approximately 70 percent of patients with MS, although these findings have never been published. Recently Gillson found severe hypochlorhydria or achlorhydria in 9 of 14 (64%) patients with MS, also using the Heidelberg capsule. Fasting plasma essential amino acid levels were also measured in 12 unselected patients with MS, prior to their starting histamine treatment. Ten of 12 had at least five ab-

Table 1. Gastric Acid Secretary Status and Amino Acid Status in MS Patients.

Patient Initials	Impaired gastric acidity	Abnormal levels of more than 5 essential amino acids	Low or borderline plasma Histidine level
BG	✓	✓	✓
JR		✓	✓
JW	✓	✓	✓
GW	✓	✓	Low normal
LM	✓	✓	✓
JM	Not tested	✓	✓

trols. A total of 24 amino acids were measured in the study, and the levels of other amino acids were “more or less the same in the two groups” according to the abstract, although histidine was not mentioned specifically.

Overall, the present work supports published abnormalities of amino acid levels in general in MS patients. Our work and one previous study also show decreased histidine levels in MS; findings from two previous studies are

normal values (out of 10 measured).

A small number of MS patients at our (JW/GG) clinic recently had both a Heidelberg assay as well as measurement of fasting plasma essential amino acids, including histidine. Six of six patients had low values on six or more of the 10 essential amino acids measured. Five of the six had histidine levels very close to the low end of the normal range, and five of six patients were achlorhydric or severely hypochlorhydric. Five of these six patients have had a favorable response to exogenous histamine therapy. These results are summarized in Table 1.

Little has been written about plasma amino acid levels in MS. Oriente¹⁰⁰ noted significantly decreased histidine, methionine, and valine levels in nine patients with progressive systemic sclerosis (equivalent to MS in the Russian literature) compared to 15 controls. A study by Ivanokov¹⁰¹ on 45 MS patients reported “marked deaminoacidemia tending to hypoaminoacidemia.” The abstract did not elaborate regarding which amino acids were measured. Quereshi¹⁰² noted decreased levels of methionine, valine, phenylalanine, and lysine in 12 patients with MS versus 12 con-

equivocal. Nyhan reported five cases of hypohistidinemia in males without MS. Interestingly, all had CNS abnormalities.¹⁰³

Evidence of malabsorption of other nutrients, such as zinc¹⁰⁴ and copper^{104,105} has also been reported in MS. Both zinc and copper deficiency have been tied to histidine depletion in dogs,¹⁰⁶ and copper deficiency has been linked to demyelination, as mentioned earlier. Copper is also a mast cell stabilizer.^{107,108} A possible connection between CNS histidine deficiency, copper deficiency, mast cell degranulation, and demyelination might, therefore, be worth investigating.

Proposed Connections between Histamine, Histidine, and the Pathophysiology of MS

With the foregoing background information in mind, the framework outlined in Figure 2 is proposed.

A central feature is the establishment of a systemic depletion of histidine on the basis of impaired gut function, as well as possible additional histidine depletion due to ongoing or intermittent episodes of CNS mast cell

degranulation. Mast cell mediators are presumed to be involved in promulgation of demyelination, as discussed earlier. Demyelination is assumed to underlie some of the motor symptoms seen in MS, in accord with accepted thinking.

Histidine is needed for histamine synthesis (both systemically and in the CNS) for mast cells, and histaminergic neurons. Histidine deficiency may, therefore, lead to impairment of histaminergic pathways in the CNS via diminished release of histamine in synaptic clefts of histaminergic synapses. We postulate that some of the symptoms of MS, such as fatigue, impaired balance, and poor heat tolerance might be due to this impairment.

The Possible Role of Carnosine

Carnosine is a histidine-alanine dipeptide found primarily in skeletal muscle, cardiac tissue, and the brain.¹⁰⁹ A murine study showed carnosine can be absorbed intact from the gut.¹¹⁰ If the same is true in humans, carnosine might be subject to the same factors putatively impairing histidine assimilation in MS. A carnosine deficiency might then contribute to an existing histidine deficit, since carnosine might provide a rapidly mobilizable source of histidine in times of high histamine turnover.¹¹¹ Both Fitzpatrick¹¹² and Flancbaum¹¹³ demonstrated increased conversion of carnosine to histamine in various tissues of animals under stress. Carnosine may also act as a histidine reservoir for normal metabolic demands. Prolonged or episodic inflammatory episodes in MS might, therefore, eventually deplete carnosine muscle stores, which would not be replenished by uptake from the gut. Deficiency of carnosine might also make the CNS more susceptible to damage from inflammation, in light of its status as a lipophilic antioxidant.¹¹⁴⁻¹¹⁶

Conversion of carnosine to histidine is carried out by the enzyme carnosinase, and decreased carnosinase activity has been

demonstrated in MS.¹¹⁷ This may be due to subtle thyroid impairment, since decreased serum carnosinase activity is also seen in hypothyroidism.¹¹⁸ Kiessling¹¹⁹ observed elevated T4 levels with reduced T3 levels in MS patients, suggesting possible impaired conversion of T4 to T3.

The Possible Role of Thiamine

Histidine residues are known to be critical sites in thiamine-binding proteins, such as thiamine pyrophosphokinase, the enzyme that generates thiamine pyrophosphate from thiamine and ATP.¹²⁰ Histidine residues are also critical residues in other proteins known to bind TPP or thiamine itself.^{121,122} Histidine deficiency might therefore result in decreased TPP synthesis or decreased transport of TPP or thiamine to relevant sites such as the brain.

As mentioned, thiamine is involved in maintaining the stability of the blood-brain barrier and also in myelin formation — two central aspects of MS pathophysiology. Thiamine deficiency can also feed back to impair gut function in rats, as shown by Khramtsov¹²³ who demonstrated impaired gastric secretory function in response to histamine in thiamine-deficient animals.

Given the dependence of thiamine binding proteins on histidine residues, abnormal thiamine status might develop in MS as a consequence of histidine deficiency, and once established, may reinforce the histidine deficiency. A thiamine deficiency would then promote demyelination as discussed above; alternatively, subclinical thiamine deficiency might be a factor which tips a predisposed individual closer to acute MS or relapse.

A Possible Viral Connection

The notion that a histamine deficit exists in the histaminergic neurons of some patients with MS due to a slow viral attack has been proposed by DeLack.¹²⁴ If such a viral attack occurs, it might be centered on the

hypothalamus, since this is where histaminergic neurons originate. Since these histaminergic neurons regulate basic physiologic functions, one might expect the course of MS to be more aggressive, with more pronounced disruption of “life support” functions. There is at least one report in the literature¹²⁵ of wasting and a rapidly fatal course in MS, with lateral hypothalamic lesions, but such presentations are rare. Immunologic assays directed toward detection of such a viral attack on histaminergic neurons might help to determine the extent and selectivity of a possible viral attack on histaminergic neurons.

Detailed Discussion of Potential Mechanisms of Action of Exogenous Histamine in the Proposed Framework

Symptom alleviation in patients who have benefited from exogenous histamine has at times been rapid — hours to several days.¹ One inference, which might be drawn from this observation, is that the effects are due to the direct action of histamine once a threshold concentration has been achieved in the target tissue. If the primary mechanism involved is an indirect action such as modulation of the immune response, stimulation of repair of damaged tissues, or improved nutritional status, one might expect a longer timescale of action (similar, for example, to the response of rheumatoid arthritis to disease-modifying agents). Some users of exogenous histamine report they continue to improve over a time span of weeks to months. This suggests that mechanisms with longer timescales of action might also be operative in some cases.

Effect of Exogenous Histamine on Gut Dysfunction

Local release of histamine from the ECL cells of the gastric mucosa stimulates

release of acid by an H₂ receptor-mediated mechanism. Histamine also stimulates the release of pancreatic secretions via H₁ receptor stimulation.¹²⁶⁻¹²⁸ If there is a local deficiency of histamine synthesis in the ECL cells, exogenous histamine may stimulate gastric and pancreatic function, improve protein hydrolysis, and eventually replenish histidine and carnosine levels. Improved gastric acid secretory function might improve B₁₂ and thiamine absorption, with possible beneficial effects on myelination. Absorption of various other nutrients important for maintaining myelin (such as copper) should also improve due to improved gastric acidity.

Histaminergic fibers have been noted in the submucous ganglion cell layer of the ileum of the rat and the guinea pig.¹²⁹ If such fibers are also present in the human ENS, they may be involved in the regulation of peristalsis, as well as the regulation of secretory functions through the action of histamine either on histamine receptors or serotonin receptors (cross-talk between histamine and serotonin receptors is discussed in reference 20). Diminished histamine synthesis in these postulated enteric histaminergic pathways might therefore impair secretory and peristaltic functions. Conversely, exogenous histamine might improve the function of enteric histaminergic pathways by directly increasing the concentration of histamine in relevant synapses, or indirectly, by stimulating gastric acid production and absorption of histidine, and eventually improving endogenous histamine production.

At this juncture, it is not clear whether gut dysfunction is a primary or secondary feature of MS. Primary failure mechanisms might include a cell-mediated attack on the ENS which parallels that experienced by the CNS, or a viral attack on intestinal mucosa, pancreatic tissue, and/or neuronal tissue. Secondary mechanisms might include a disruption of CNS vagal efferents (due to the effects of MS in the brain), which are involved

in the control of gut function. One of the authors (GG) has measured achlorhydria in two patients within months of first onset of neurologic symptoms, supporting the notion that gut dysfunction might precede development of neurologic symptoms.

The concept that pathological processes affecting the CNS might be mirrored in the ENS is discussed by Gershon⁹⁶ and is supported by research on patients with Parkinson's disease. Singaram¹³⁰ observed depletion of colonic dopaminergic neuron populations in 9 of 11 Parkinson's patients, and decreased levels of dopamine in muscularis externa specimens in four of four patients versus zero of four controls. Wakabayashi¹³¹ also reported the presence of Lewy bodies — markers of neuronal degeneration characteristic of Parkinson's disease — in the ENS (myenteric and submucosal plexuses from the upper esophagus to the rectum) of 28 of 30 patients with Parkinson's disease.

Measles viral antigens have been noted in jejunal biopsies of MS patients,³² supporting the notion that direct viral attack on the gut might be a factor in some patients.

Central vagal outflow is important for gastric acid secretion,¹³² and various investigators have studied the central effect of histamine on vagal tone. Both inhibitory and stimulatory effects, depending on histamine concentration, have been observed.^{133,134} Vagal tone is also influenced by other factors, including thyrotropin releasing hormone¹³⁵ and inflammatory mediators such as IL-1 β .¹³⁶ Failure of central vagal outflow to the stomach might explain reduced gastric acid secretion in MS, but it is unclear how this might arise, or whether an alteration in central histaminergic activity is even involved.

A reasonable body of evidence supports the existence of impaired digestion and uptake of nutrients in MS, but the mechanisms are not well understood at this time. This is an important area that needs additional study, since similar gut dysfunction

may be associated with other chronic diseases. For example, Gerber¹³⁷ reported on 26 patients with active rheumatoid arthritis who had significantly lower serum histidine levels compared to controls. Also, histidine has been shown to be an important residue for binding and uptake of other nutrients, including biotin¹³⁸ and folate.¹³⁹ Histidine-deficient dogs also had lower whole blood concentrations of zinc and copper,¹⁰⁶ as mentioned earlier.

Histidine levels and gastric acid secretory capacity data are needed from a larger number of MS patients. Data on thiamine and carnosine status would also help to confirm or refute some of our hypotheses. It remains to be seen whether the MS patients studied via Heidelberg capsule will recover gastric acid secretory capability with long-term transdermal histamine supplementation. Immunohistochemical studies of pathology specimens obtained from MS patients who undergo intestinal surgery could be fruitful to delineate the role of histaminergic fibers in the human ENS, and whether the ENS in general exhibits demyelination or other degeneration in MS.

Effect of Exogenous Histamine on the Proposed Histaminergic Neuronal Histamine Deficit

Exogenous histamine supplementation might directly increase the concentration of histamine in CNS synaptic clefts, as well as indirectly through increased synthesis of histamine (due to increased availability of histidine through effects on the gut). Either way, increased synaptic cleft histamine might account for improvements in symptoms such as fatigue, balance impairment, and heat intolerance. This is envisioned to be rapid-acting if direct, slower if indirect.

It might reasonably be asked if any decrease in the CSF histamine level is seen in MS patients (assuming CSF is representative of the environment within the synaptic clefts). CSF histamine concentrations have been

measured by various researchers, with inconsistent findings. Suojaranta-Ylinen¹⁴⁰ measured CSF histamine levels in normal patients and found them to be on the order of 60 ± 40 pg/ml. Kiviranta¹⁴¹ found histamine levels in the CSF of 21 normal children to be 40 ± 8 pg/ml. Rozniecki¹⁴² found histamine concentrations in the CSF of 55 patients with MS to be $40 \pm 40-80$ pg/ml and saw no difference, compared to 39 controls with other neurologic disease.

In the same study, Rozniecki also saw no difference in CSF tele-methylhistamine (the CNS metabolite of histamine) levels between controls and MS patients. The level of tryptase, a mast cell mediator, was however, found to be significantly higher in MS patients, compared to controls ($p < 0.002$). Rozniecki's data, therefore, reflect evidence of mast cell degranulation, but not histamine elevation.

The findings of both Tuomisto¹⁴³ and Molnar¹⁴⁴ are markedly in contrast, and show a significant elevation of CSF histamine levels in patients with MS compared to controls. Elevated levels were seen both in patients with relapsing-remitting and progressive MS. Control levels of histamine were roughly 10-fold higher in Tuomisto's study (440 pg/ml) and 50-fold higher in Molnar's study (2200 pg/ml) compared to the other three studies (40 pg/ml). The data from the three most recent papers (<10 years old)¹⁴⁰⁻¹⁴² are all in agreement. The two earlier papers are older, and it is difficult to say how much of the disparity between the two groups of results is due to differences in analytical methods. It is also possible that CSF histamine levels are simply not representative of histamine status in brain tissue, as was proposed by Rozniecki.¹⁴² For these reasons, no firm conclusions may be drawn from the CSF histamine data.

The only other data with even a tangential connection to synaptic cleft histamine levels was reported by Iarosh⁹⁸ who measured whole blood histamine levels in controls and MS patients as a function of

disease duration. He reported an elevation of histamine levels compared to controls in patients who had MS less than five years, with the greatest elevation seen less than two years after disease onset. Patients who had MS greater than five years were found to have lower blood histamine levels than controls. Patients with gastric ulcerations (shown by concurrent endoscopic studies) had the lowest histamine levels.

Once again, it is difficult to interpret this data. Whole blood histamine levels vary with both sex and age, with females having higher histamine levels at any given age, and levels in both sexes declining with age.¹⁴⁵ Unless control patients were age and sex matched, there is potential for significant bias in the Iarosh study. Unfortunately, the age and sex stratifications of the study and control groups were not provided in the article.

Histamine is intimately involved in various aspects of reproductive physiology, including uterine contractility (guinea pig),¹⁴⁶ estrogen and progesterone secretion by human ovarian granulosa cells,¹⁴⁷ and ovulation (hamster).¹⁴⁸ Thus, an increased need for histidine and histamine for orchestration of reproductive function in women seems reasonable. It is possible that the whole blood histamine level reflects the body's histidine pool, since basophils, eosinophils, and platelets all either synthesize or store histamine. This might explain why young women have the highest whole blood histamine levels, and why these levels peak in the childbearing years.¹⁴⁵ The blood level may merely reflect the need for higher circulating histidine levels (to support histamine's role in reproduction).

From the foregoing, any process which impairs CNS availability of histidine might tend to affect women of reproductive age more than men, since the body might tend to divert available histidine toward reproductive endeavors, and short-change the CNS. Because the incidence of MS is highest in women of

reproductive age, another tentative link between MS and histidine/histamine might be proposed; i.e., those with a greater systemic need for histidine/histamine might be more susceptible to MS.

It appears the available evidence from CSF and blood does not conclusively support or refute the concept of a deficit of histamine in histaminergic neurons or synapses. Also, it remains to be explained how a histamine deficit could even arise in the face of mast cell degranulation, with release of presumably copious amounts of non-synaptic histamine. Non-synaptically released histamine should still be able to diffuse into the synaptic clefts and “make up” for any intrinsic neuronal synthetic deficit. This seems logical, but in the face of ongoing or recurrent inflammatory episodes, we postulate a depletion of mast cell histamine (due to a lack of histidine precursor) but not other inflammatory mediators. Degranulation would still occur, but non-synaptic histamine would no longer be available to drift into the histamine synapses. In the light of this idea, perhaps Rozniecki’s data can be interpreted in a different way; i.e., no histamine was seen in the CSF because mast cells were no longer releasing it (although they were still degranulating as evidenced by the tryptase data).

Accepting that a histaminergic synaptic histamine deficit may exist at some point after the onset of MS, how might such a deficit cause symptoms, such as difficulty maintaining balance when walking, fatigue, and heat intolerance? How are histaminergic neurons involved in these aspects of function?

Hypothalamic histaminergic neurons project to the human cerebellum as shown by Panula,¹⁰ and also to the vestibular nuclei in the cat brainstem.¹⁶ Both areas are important for maintaining balance; the cerebellum for coordination of motor impulses, and the vestibular nuclei for processing sensory afferents from the vestibular apparatus in the inner ear. A left-right imbalance in electrical or caloric

stimulation of the middle ear (effectively stimulating the inner ear) resulted in increased firing of hypothalamic histaminergic neurons in rats.¹⁷ Decreased release of histamine by relevant histaminergic neurons in MS might, therefore, result in problems with balance. None of these studies determined the relevant receptor type, although H1 receptor blockade is a common pharmacological intervention for vertigo arising from vestibular disorders.

Fatigue is a common, though difficult to define, symptom in MS.¹⁴⁹ Many MS patients describe their fatigue as a lack of motivation or lack of interest, as well as actual sleepiness or difficulty staying awake. Although the cause of “fatigue” is undoubtedly multifactorial, many users of exogenous histamine report a substantial improvement in this symptom in the personal experience of the principal author. This may be partly attributable to the action of exogenous histamine at H1 receptors, since H1 receptor stimulation appears to have an animating effect on behavior. H1-receptor-deficient rats displayed significantly less interest in their environment and less activity on an exercise wheel,¹² and H1 receptor blocking agents are also common ingredients in non-prescription sleep aids.

Heat intolerance is a common symptom in MS.¹⁵⁰ The accepted explanation for worsening of MS symptoms upon heat exposure is conduction block of demyelinated nerve fibers, although at least one author has proposed the etiology of heat intolerance is multifactorial, since conduction block does not explain various anomalous or paradoxical responses.¹⁵¹ Therefore, it is worthwhile to examine the potential role histamine may play in the response to warming.

Intracerebroventricular injection of histamine lowered rectal temperature in rats; this was shown to be mediated by both H1 and H2 receptors.¹⁵ We postulate that histamine release by hypothalamic histaminergic neurons might be part of the response to warming in humans also. Mounting the proper response

to warming (such as increased skin vasodilation) likely takes precedence over other CNS uses of histamine, since brain temperature is a vital physiologic parameter. In MS patients, warming might therefore consume available histidine to stabilize brain temperature at the expense of other histamine-requiring neurologic processes, including those responsible for maintaining alertness, and may thus result in the “wilting” or fatigue experienced with warming. Once again, exogenous histamine may mitigate this effect by directly increasing the amount of histamine available; this should manifest over a short time period.

Histamine is also well recognized as a peripheral vasodilator.¹⁵² Increased blood flow to the skin with exogenous histamine supplementation should improve heat tolerance by improving the ability to release excess heat through the skin.

The Effect of Exogenous Histamine on the Electrical Properties of Demyelinated Nerve Fibers

The foregoing has focused on suboptimal functioning of otherwise undamaged histamine-dependent neural pathways, which might be improved by exogenous replacement of histamine. The majority of neurologic deficits in MS however, are thought to be due to faulty conduction by demyelinated fibers, although there is some evidence that reversible secondary factors are important.¹⁵³ Rapid-onset beneficial effects of histamine supplementation might be expected if histamine can enhance the conduction of demyelinated fibers; moreover, this should be a reversible phenomenon, with effects being observed only when sufficient histamine is present in the tissue around the demyelinated axons.

A “booster” effect of this type has been observed with 4-aminopyridine (4AP), which is known to modify conduction in demyelinated nerves by blocking potassium channels.¹⁵⁴ In two animal studies 4AP also

restored conduction in blocked, demyelinated nerves.^{155,156} Two controlled trials^{157,158} have demonstrated the ability of 4AP to reversibly improve symptoms of MS. One researcher commented that effects usually manifested within 60 minutes of oral administration, and reversed gradually over four to seven hours.¹⁵⁷ The clinical utility of 4AP is somewhat compromised by side-effects, including dizziness, paresthesias, and seizure in higher doses.¹⁵⁸

Histamine has been shown to facilitate nerve conduction in the same fashion as 4AP; i.e., by blocking a slow polarizing potassium current. This effect has been observed in human cortical neurons,¹⁵⁹ rat cholinergic neurons,¹⁶⁰ rat supraoptic nucleus neurons,¹⁶¹ and ferret vagal afferent neurons.¹⁶² Histamine was also shown to depolarize neurons by inhibition of a slow post-spike hyper-polarizing potassium aftercurrent.^{162,163} Both effects were mediated through H1 receptor activation.

There have been reports of synergistic effects of histamine and 4AP. Histamine-induced vasoconstriction was augmented in rabbit ear arteries by 4AP-stimulated release of glutamate from hippocampal neurons.^{164,165} Co-administration of histamine and 4AP might, therefore, prove beneficial for enhancing nerve conduction in demyelinated fibers.

If this mechanism is operative to any significant extent, then treatment with H1 blockers might be expected to abolish its effect. A study of the effects of H1 blockers on the response of MS patients to exogenous histamine should clarify the relative importance of this mechanism.

Effect of Exogenous Histamine on Immune Function

Histamine can suppress its own release by inhibiting mast cell degranulation. Its possible role as a modulator of the severity of inflammation was proposed by Bourne.¹⁶⁶ Exogenous histamine might, therefore, serve to

slow or halt the inflammatory spiral provoked by the release of other CNS mast cell inflammatory mediators, taking over the role of endogenous CNS mast cell histamine if its synthesis and release is impaired by decreased availability of histidine. This effect could operate over both short and long timescales.

It is generally recognized that histamine can exert bivalent effects on the immune system, with stimulatory effects generally mediated by H1 receptors and suppressive effects generally mediated by H2 receptor stimulation.²¹ For example, mouse H1 receptor-deficient T cells demonstrated impaired proliferative response to antigens in one study,¹⁶⁷ and subcutaneous histamine administration (4 mg/day) plus an oral H2 blocker (thereby stimulating H1 receptors) resulted in prolongation of survival in patients with metastatic melanoma.³ This is the same dose range currently being used in MS (3-6 mg/day), suggesting small amounts of histamine seem to exert pronounced effects on immune function. Inhibition of IL-2 production in mitogen-stimulated human monocytes was shown to be mediated by H2 receptors.¹⁶⁸

Sometimes the inhibitory effects of H2 receptor stimulation can enhance the overall immune response. In one report, the tumor cell-killing ability of natural killer (NK) cells was enhanced by the addition of histamine.¹⁶⁹ The effect was shown to be H2 receptor-mediated and involved suppression of the release of NK cell-toxic reactive oxygen intermediates from intratumoral monocytes.

Exogenous histamine may also be acting to suppress the overactive Th1 cell-mediated immune response seen in MS and boost the underactive Th2 response. Two researchers^{170,171} have demonstrated that histamine (H2 receptor-mediated) can suppress the production of IL-12. Elenkov also showed histamine stimulates the production of IL-10, once again by H2 receptor stimulation.¹⁷⁰ As discussed earlier, down-regulation of IL-12 and up-regulation of IL-10 is thought to be beneficial in

MS. Exogenous histamine might exert an overall favorable effect on the immune system in MS by this mechanism, although this has not been studied. Note also that histamine may exert the same effect on Th1/Th2 balance as copolymer 1;⁴⁰ this suggests the two might be used simultaneously for a synergistic effect.

The concept that histamine release may provide negative feedback on inflammation mediated by mast cells was mentioned above. The dose-dependent inhibition of antigen-induced release of histamine from human leukocytes by exogenous histamine reported by Bourne was shown to be H2 receptor mediated.¹⁷² Histamine has been shown to exert the same effect on CNS mast cells via H3 receptor stimulation.²² Therefore, there is reasonable support for the view that exogenous histamine might exert a braking effect on CNS inflammation in MS, possibly by balancing Th1/Th2 activity and by stabilizing mast cells.

Caffeine is included in the transdermal histamine patch utilized in the Part One study previously published in this journal¹ and may synergize with histamine in several ways. Mast cell degranulation involves several processes: an influx of Ca²⁺ ions to the cell, a transient decrease in the intracellular concentration of cAMP,¹⁷³ and a transient increase in the activation of protein kinase C (PKC).¹⁷⁴ Phosphodiesterase inhibitors, such as theophylline¹⁷³ and caffeine¹⁷⁵ are able to stabilize mast cells against degranulation by preventing a drop in intracellular cAMP levels and preventing activation of PKC. Therefore, caffeine would synergize with histamine in this regard.

Most H2 receptors have cAMP as a second messenger; i.e., stimulation of an H2 receptor by histamine results in the transient increase in intracellular cAMP. Co-administration of a phosphodiesterase inhibitor with histamine would then serve to amplify the "message" delivered by histamine, by prolonging the elevation of cAMP. Thus, cAMP-specific phosphodiesterase inhibition would seem to be a therapeutic strategy worth

investigating. In fact, phosphodiesterase Type IV inhibitors have been proposed as a treatment for MS¹⁷⁶ and have been successfully employed in the treatment of EAE.¹⁷⁷

Recent work has shown phosphodiesterase inhibitors exhibit Th1-suppressing and Th2-enhancing effects on human leukocytes¹⁷⁸ and in MS patients;¹⁷⁹ as mentioned, histamine also exerts these effects. If caffeine acts similarly to other phosphodiesterase inhibitors, this is yet another point in favor of the concurrent use of histamine and caffeine. Caffeine may reinforce the actions of histamine and in parallel, may produce some of the same immunomodulating effects as histamine.

Since the CNS damage in MS is presumed to be due to the influx of autoreactive T cells, permeability of the blood-brain barrier (BBB) is an important factor to consider when discussing aspects of the immune response in MS. Both Sharma⁴⁹ and Gulati⁹² have shown histamine increases BBB permeability through H2 receptor stimulation, which suggests exogenous histamine should worsen MS symptoms by increasing BBB permeability. It may well be that exogenous histamine does increase BBB permeability (which could easily be proven with an MRI tracer study), but potential negative sequelae of this permeability increase are over-ridden by other beneficial effects.

The Effect of Exogenous Histamine on Myelination

Exogenous histamine may exert a positive influence on remyelination by direct and indirect means. Exogenous histamine might directly stimulate remyelination by enhancing migration of oligodendrocyte progenitor cells into areas of inflammation, by inducing and/or facilitating the development of mature oligodendrocytes, and by stimulating myelin formation by oligodendrocytes. Although stimulation of H2 receptors elevates intracellular cAMP, and elevation of intracellular cAMP has

been linked to induction of oligodendrocyte differentiation and myelin synthesis, no studies directly connect histamine to myelin formation through elaboration of cAMP.

Exogenous histamine may indirectly spur remyelination by its effect on other factors tied to myelination, including T3 and vitamin B12. For example, histamine is known to trigger the release of thyrotrophin releasing hormone,¹⁸⁰ and mouse thyroid has been shown to bear H2 receptors, which stimulate thyroid hormone secretion.¹⁸¹ Histamine may exert a similar stimulating effect on the human thyroid gland, so exogenous histamine may assist remyelination through stimulation of thyroid function. Exogenous histamine might enhance uptake of B12 in particular, and other nutrients in general, and this may impact myelination over a longer time period. Note also that improved thyroid function might increase carnosinase activity.

The Effect of Exogenous Histamine on Oxygenation of Cerebral Tissues

Decreased cerebral blood flow (CBF) and decreased oxygen metabolism are well recognized in MS, as demonstrated by early studies employing radio-labeled xenon gas¹⁸² and later studies employing MR¹⁸³ and PET¹⁸⁴ imaging. Both white and grey matter exhibit decreased cerebral blood flow. Lycke found decreased CBF correlated to neurologic disability, cognitive function, and visual performance.¹⁸⁵ Sun demonstrated cerebral hypometabolism (a measure of oxygen consumption) was correlated with the number of relapses.¹⁸⁴ Lycke also demonstrated that patients with progressive MS had a greater decrease in grey matter CBF than relapsing-remitting patients.¹⁸⁵

Recent data concerning the effect of histamine on CBF generally agree. Various researchers have reported that histamine stimulates vasodilation by an H2 receptor-mediated mechanism, and vasoconstriction by an H1

receptor-mediated effect. The observed effect (vasodilation or vasoconstriction) is concentration dependent,¹⁸⁶⁻¹⁸⁸ and also depends on the anatomic location of the vessel.¹⁸⁸ No recent studies measuring the net effect of low-dose systemic histamine administration on CBF were found, but older studies may shed some light on the question.

In the January 1951 issue of *Postgraduate Medicine*, Horton stated, "The results of a recent study indicate that histamine causes a greater increase in blood flow to the CNS than any other drug known. Photographic records of the pulsations of the human brain indicated that the subcutaneous administration of 0.25 mg of histamine base produced a 725 percent increase in the amplitude of the pulsations of the brain."¹⁸⁹ Later in the same article, Horton also reported that during an exploratory craniotomy, he observed the following effect of the intravenous administration of histamine: "The cortex became fiery red and vessels which usually were hardly visible became prominent."

These data support the notion that exogenous histamine increases CBF and oxygenation. Increased oxygen delivery could lessen fatigue, increase alertness, and increase cognitive performance. Over the long term, continued augmentation of CBF could also exert beneficial effects through stimulation of repair by enhanced delivery of oxygen and nutrients.

Conclusion

A framework has been proposed in which histamine supplementation might alleviate some of the symptoms of MS. Central to this framework is the notion of a systemic histidine deficiency, which limits the ability of the body to synthesize requisite amounts of histamine at sites such as the CNS and the gastric mucosa. We have reported preliminary findings which support the existence of a histidine deficiency in some patients with MS, and correlate it with impaired or absent gastric acid production as measured by the Heidelberg

capsule. The proposed histidine deficiency might be brought about by a failure of gastric and pancreatic exocrine function, but it is unclear whether impairment of gut function might be a primary or secondary feature of MS, and what might initiate the impairment. CNS mast cell degranulation may also contribute to a shortage of histidine.

Histamine supplementation may impact this state of affairs by directly augmenting the concentration of histamine in relevant synaptic clefts, by suppressing CNS mast cell degranulation, or by improving protein digestion and histidine status. Exogenous histamine might also augment cerebral blood flow, improve the conductivity of demyelinated nerve fibers, or help balance Th1 and Th2 immune functions. The timescales over which these mechanisms might act are roughly consistent with observed timescales of improvement.

As pointed out in the introduction, many of the ideas discussed in this paper are speculative. A host of unanswered questions remain, and we have proposed various ways some of these questions might be addressed. Whole brain n-acetylaspartate (NAA) assay via MRI has recently been proposed as a marker of disease progression.¹⁹⁰ This technique might prove to be particularly useful for studying mast cell degranulation and the role of histamine in MS, if brain mast cells contain NAA as do peritoneal mast cells.¹⁹¹ We hope histamine will continue to show promise for MS patients, and that the curiosity of other MS and histamine researchers will be stimulated by some of the ideas presented in this paper.

Secondary deficits of nutrients such as histidine, thiamine, carnosine, and copper may feed back and amplify the damage done by the primary disease process in MS. We believe broad-spectrum nutritional support along with efforts to improve gastric acidity and protein digestion are strongly indicated, in conjunction with whatever other treatment modalities are used for this disease. We also believe significant effort should be put into untangling

the interlocking ways in which secondary nutritional deficits sustain and intensify MS and other chronic disease processes.

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References

- Gillson G, Wright JV, DeLack E, Ballasiotes G. Clinical experience with transdermal histamine in multiple sclerosis and a proposed theoretical basis for its efficacy. Part I. *Alt Med Rev* 1999;4:424-428.
- King WP. The use of low-dose histamine therapy in otolaryngology. *Ear Nose Throat J* 1999;366-368.
- Burtin C, Noirot C, Scheinmann P, et al. Clinical improvement in advanced cancer disease after treatment combining histamine and H2-antihistaminics (ranitidine or cimetidine). *Eur J Cancer Clin Oncol* 1988;24:161-167.
- Horner M, Helle J, Schurmann FW. The distribution of histamine-immunoreactive neurons in the ventral nerve cord of the cricket, *Gryllus bimaculatus*. *Cell Tissue Res* 1996;286:393-405.
- Battelle BA, Calman BG, Hart MK. Cellular distributions and functions of histamine, octopamine, and serotonin in the peripheral visual system, brain, and circumesophageal ring of the horseshoe crab *Limulus polyphemus*. *Microsc Res Tech* 1999;44:70-80.
- Weinreich D. Multiple sites of histamine storage in superior cervical ganglia. *Exp Neurol* 1985;90:36-43.
- Panula P, Flugge G, Fuchs E, et al. Histamine-immunoreactive nerve fibers in the mammalian spinal cord. *Brain Res* 1989;484:234-239.
- Airaksinen M, Paetau A, Paljarvi L, et al. Histamine neurons in human hypothalamus; anatomy in normal and Alzheimer diseased brains. *Neuroscience* 1991;44:465-481.
- Wada H, Inagaki N, Yamatodani A. Histaminergic neuron system in the brain: distribution and possible functions. *Brain Res Bull* 1991;27:367-370.
- Panula P, Takagi H, Inagaki N, et al. Histamine-containing nerve fibers innervate human cerebellum. *Neurosci Lett* 1993;160:53-56.
- Tuomisto L, Eriksson L. Histamine and histamine-N-methyltransferase in the goat brain. *Agents Actions* 1982;12:142-145.
- Yanai K, Son LZ, Endou M, et al. Behavioural characterization and amounts of brain monoamines and their metabolites in mice lacking histamine H1 receptors. *Neuroscience* 1998;87:479-487.
- Lecklin A, Etu-Seppala P, Stark H, Tuomisto L. Effects of intracerebroventricularly infused histamine and selective H1, H2 and H3 agonists on food and water intake and urine flow in Wistar rats. *Brain Res* 1998;793:279-288.
- Kjaer A, Knigge U, Rouleau A, et al. Dehydration-induced release of vasopressin involves activation of hypothalamic histaminergic neurons. *Endocrinology* 1994;135:675-681.
- Chen Z, Sugimoto Y, Kamei C. Effects of intracerebroventricular injection of histamine and its related compounds on rectal temperature in mice. *Methods Find Exp Clin Pharmacol* 1995;17:669-675.
- Tighilet B, Lacour M. Distribution of histaminergic axonal fibres in the vestibular nuclei of the cat. *Neuroreport* 1996;7:873-878.
- Horii A, Takeda N, Matsunaga T, et al. Effect of unilateral vestibular stimulation on histamine release from the hypothalamus of rats in vivo. *J Neurophysiol* 1993;70:1822-1826.
- Knigge U, Warberg J. Neuroendocrine functions of histamine. *Agents Actions Suppl* 1991;33:29-53.
- Knigge U, Willems E, Kjaer A, et al. Histaminergic and catecholaminergic interactions in the central regulation of vasopressin and oxytocin secretion. *Endocrinology* 1999;140:3713-3719.
- Mitsuhashi M, Payan D. Functional diversity of histamine and histamine receptors. *J Invest Dermatol* 1992;98:8S-11S.
- Medina M, Quesada AR, de Castro I, Sanchez-Jimenez F. Histamine, polyamines and cancer. *Biochem Pharmacol* 1999;57:1341-1344.
- Rozniecki JJ, Letourneau R, Sugiultzoglou M, et al. Differential effect of histamine 3 receptor-active agents on brain, but not peritoneal, mast cell activation. *J Pharmacol Exp Ther* 1999;290:1427-1435.

23. Maeyama K, Watanabe T, Yamatodani A, et al. Effect of alpha-fluoromethylhistidine on the histamine content of the brain of W/W^v mice devoid of mast cells: turnover of brain histamine. *J Neurochem* 1983;41:128-134.
24. Yamatodani A, Maeyama K, Watanabe T, et al. The contents of histamine in various tissues of mutant mice deficient in mast cells: clear evidence for the presence of non-mast cell histamine. *Biochem Pharmacol* 1982;31:305-309.
25. Letourneau R, Pang X, Sant GR, Theoharides TC. Intragranular activation of bladder mast cells and their association with nerve processes in interstitial cystitis. *Br J Urol* 1996;77:41-54.
26. Tamura K, Kogo H. Granulocyte-macrophage colony-stimulating factor enhances interleukin-1-beta stimulated histamine release in the preovulatory rat ovary. *Eur J Pharmacol* 1999;373:207-213.
27. Mayerhofer A, Bartke A, Amador AG, Began T. Histamine affects testicular steroid production in the golden hamster. *Endocrinology* 1989;125:2212-2214.
28. Merrill JE, Strom SR, Ellison GW, Myers LW. In vitro study of mediators of inflammation in multiple sclerosis. *J Clin Immunol* 1989;9:84-96.
29. Ewing C, Bernard CC. Insights into the aetiology and pathogenesis of multiple sclerosis. *Immunol Cell Biol* 1998;76:47-54.
30. Cook AW, Pertschuk L, Gupta J, Kim D. Jejeunal virus antigen in multiple sclerosis and amyotrophic lateral sclerosis. *Adv Exp Med Biol* 1978;100:627-632.
31. Soldan SS, Berti R, Salem N, et al. Association of human herpes virus 6 (HHV-6) with multiple sclerosis; Increased IgM response to HHV-6 early antigen and detection of serum HHV-6 DNA. *Nature Med* 1997;3:1394-1397.
32. Kempe CH, Takabayashi K, Miyamoto H, et al. Elevated cerebrospinal fluid vaccinia antibodies in multiple sclerosis. *Arch Neurol* 1973;28:278-279.
33. Clerici M, Fusi ML, Caputo D, et al. Immune response to antigens of human endogenous retroviruses in patients with acute or stable multiple sclerosis. *J Neuroimmunol* 1999;99:173-182.
34. Garson JA, Tuke PW, Giraud P, et al. Detection of virion-associated MSR-V-RNA in serum of patients with multiple sclerosis. *Lancet* 1998;351:33.
35. Wilder RW. Hormones, pregnancy, and autoimmune diseases. *Ann NY Acad Sci* 1998;840:45-50.
36. Sinigaglia F, DiAmbrosio D, Panina-Bordignon P, et al. Regulation of the IL-12/IL-12R axis: a critical step in T helper cell differentiation and effector function. *Immunol Rev* 1999;170:65-72.
37. Nagelkerken L. Role of Th1 and Th2 cells in autoimmune demyelinating disease. *Braz J Med Biol Res* 1998;31:55-60.
38. Beck J, Rondot P, Catinot L, et al. Increased production of interferon gamma and tumour necrosis factor precedes clinical manifestations in multiple sclerosis: do cytokines trigger off exacerbations? *Acta Neurol Scand* 1988;78:318-323.
39. van Boxel-Dezaire AH, Hoff SC, van Oosten BW, et al. Decreased interleukin-10 and increased interleukin-12p40 mRNA are associated with disease activity and characterize different disease stages in multiple sclerosis. *Ann Neurol* 1999;45:695-703.
40. Aharoni R, Teitelbaum D, Sela M, Arnon R. Copolymer 1 induces T cells of the T helper type 2 that crossreact with myelin basic protein and suppress experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA* 1997;94:10821-10826.
41. Miller A, Shapiro S, Gershtein R, et al. Treatment of multiple sclerosis with copolymer-1 (Copaxone): implicating mechanisms of Th1 to Th2/Th3 immune-deviation. *J Neuroimmunol* 1998;92:113-121.
42. Sinigaglia F, DiAmbrosio D, Rogge L. Type 1 interferons and the Th1/Th2 paradigm. *Dev Comp Immunol* 1999;23:657-663.
43. Spadaro M, Amendolea MA, Mazzucconi MG, et al. Autoimmunity in multiple sclerosis: a study of a wide spectrum of autoantibodies. *Mult Scler* 1999;5:121-125.
44. Bastianello S, Pozzilli C, Bernardi S, et al. Serial study of gadolinium-DTPA MRI enhancement in multiple sclerosis. *Neurology* 1990;40:591-595.
45. Kermodé AG, Thompson AJ, Tofts P, et al. Breakdown of the blood-brain barrier precedes symptoms and other MRI signs of new lesions in multiple sclerosis. Pathogenetic and clinical implications. *Brain* 1990;113:1477-1489.

46. Kwon EE, Prineas JW. Blood-brain barrier abnormalities in longstanding multiple sclerosis lesions. An immunohistochemical study. *J Neuropathol Exp Neurol* 1994;53:625-636.
47. Harata N, Iwasaki Y. Evidence for early blood-brain barrier breakdown in experimental thiamine deficiency in the mouse. *Metab Brain Dis* 1995;10:159-174.
48. Ingalls TH. Clustering of multiple sclerosis in Galion, Ohio, 1982-1985. *Am J Forensic Med Pathol* 1989;10:213-215.
49. Sharma HS, Nyberg F, Cervos-Navarro J, Dey P. Histamine modulates heat stress-induced changes in blood-brain barrier permeability, cerebral blood flow, brain edema and serotonin levels: an experimental study in conscious young rats. *Neuroscience* 1992;50:445-454.
50. Wallace J. Multiple sclerosis and the blood-brain barrier: A novel approach in integrative care. *Int J Integrative Med* 1999 Sept/Oct:11-16.
51. Raine CS. Multiple sclerosis: TNF revisited, with promise. *Nature Medicine* 1995;1:211.
52. Rodriguez M, Scheithauer B. Ultrastructure of multiple sclerosis. *Ultrastruct Pathol* 1994;18:3-13.
53. Prineas JW, Barnard RO, Kwon EE, et al. Multiple sclerosis: remyelination of nascent lesions. *Ann Neurol* 1993;33:137-151.
54. Blakemore WF, Keirstead HS. The origin of remyelinating cells in the central nervous system. *J Neuroimmunol* 1999;98:69-76.
55. Franklin RJ, Blakemore WF. To what extent is oligodendrocyte progenitor migration a limiting factor in the remyelination of multiple sclerosis lesions? *Mult Scler* 1997;3:84-87.
56. Prineas JW, Kwon EE, Goldenberg PZ, et al. Multiple sclerosis. Oligodendrocyte proliferation and differentiation in fresh lesions. *Lab Invest* 1989;61:489-503.
57. Nieder C, Ataman F, Price RE, Ang KK. Radiation myelopathy: new perspective on an old problem. *Radiat Oncol Investig* 1999;7:193-203.
58. Tomic M, Torch S, Comte V, et al. Triiodothyronine has diverse and multiple stimulating effects on expression of the major myelin protein genes. *J Neurochem* 1992;59:1770-1777.
59. Baas D, Bourbeau D, Sarlieve LL, et al. Oligodendrocyte maturation and progenitor cell proliferation are independently regulated by thyroid hormone. *Glia* 1997;19:324-332.
60. Lovblad K, Ramelli G, Remonda L, et al. Retardation of myelination due to dietary vitamin B12 deficiency: cranial MRI findings. *Pediatr Radiol* 1997;27:155-158.
61. Sandyk R, Awerbuch G. Vitamin B12 and its relationship to age of onset of multiple sclerosis. *Intern J Neuroscience* 1993;71:93-99.
62. Sato-Bigbee C, Chan EL, Yu RK. Oligodendroglial cyclic AMP response element-binding protein: a member of the CREB family of transcription factors. *J Neurosci Res* 1994;38:621-628.
63. Gandelman KY, Pfeiffer SE, Carson JH. Cyclic AMP regulation of P0 glycoprotein and myelin basic protein gene expression in semi-differentiated peripheral neuroblastoma cell line D6P2T. *Development* 1989;106:389-398.
64. Lyons SA, Morell P, McCarthy K. Schwann cells exhibit P2y purinergic receptors that regulate intracellular calcium and are up-regulated by cyclic AMP analogues. *J Neurochem* 1994;63:552-560.
65. Read DH, Harrington DD. Experimentally induced thiamine deficiency in beagle dogs: pathologic changes of the central nervous system. *Am J Vet Res* 1986;47:2281-2289.
66. Buchman AL, Keen CL, Vinters HV, et al. Copper deficiency secondary to a copper transport defect: a new copper metabolic disturbance. *Metabolism* 1994;43:1462-1469.
67. Fox JM, Duppel W. The action of thiamine and its di- and triphosphates on the slow exponential decline of the ionic currents in the node of Ranvier. *Brain Res* 1975;89:287-302.
68. Castellano B, Gonzalez B, Palacios G. Cytochemical demonstration of TPPase in myelinated fibers in the central and peripheral nervous system of the rat. *Brain Res* 1989;492:203-210.
69. Bennett G, Hemming R. Ultrastructural localization of CMPase, TPPase, and NADPase activity in neurons, satellite cells, and Schwann cells in frog dorsal root ganglia. *J Histochem Cytochem* 1989;37:165-172.
70. Trapp BD, Itoyama Y, Sternberger NH, et al. Immunocytochemical localization of P0 protein in Golgi complex membranes and myelin of developing rat Schwann cells. *J Cell Biol* 1981;90:1-6.

71. Bebo BF Jr, Yong T, Orr EL, Linthicum DS. Hypothesis: a possible role for mast cells and their inflammatory mediators in the pathogenesis of autoimmune encephalomyelitis. *J Neurosci Res* 1996;45:340-348.
72. Dines KC, Powell HC. Mast cell interactions with the nervous system: relationship to mechanisms of disease. *J Neuropathol Exp Neurol* 1997;56:627-640.
73. Johnson D, Seeldrayers PA, Weiner HL. The role of mast cells in demyelination. 1. Myelin proteins are degraded by mast cell proteases and myelin basic protein and P2 can stimulate mast cell degranulation. *Brain Res* 1988;444:195-198.
74. Skaper SD, Facci L, Romanello S, Leon A. Mast cell activation causes delayed neurodegeneration in mixed hippocampal cultures via the nitric oxide pathway. *J Neurochem* 1996;66:1157-1166.
75. Dietsch GN, Hinrichs DJ. The role of mast cells in the elicitation of experimental allergic encephalomyelitis. *J Immunol* 1989;142:1476-1481.
76. Kimata M, Shichijo M, Miura T, et al. Ca²⁺ and protein kinase C signalling for histamine and sulfidoleukotrienes released from human cultured mast cells. *Biochem Biophys Res Commun* 1999;257:895-900.
77. Neu I, Mallinger J, Wildfeuer A, Mehlber L. Leukotrienes in the cerebrospinal fluid of multiple sclerosis patients. *Acta Neurol Scand* 1992;86:586-587.
78. Prosiel M, Neu I, Mallinger J, et al. Suppression of experimental autoimmune encephalomyelitis by dual cyclo-oxygenase and 5-lipoxygenase inhibition. *Acta Neurol Scand* 1989;79:223-226.
79. Dietsch GN, Hinrichs DJ. Mast cell proteases liberate stable encephalitogenic fragments from intact myelin. *Cell Immunol* 1991;135:541-548.
80. Ozenci V, Rinaldi L, Teleshova N, et al. Metalloproteinases and their tissue inhibitors in multiple sclerosis. *J Autoimmun* 1999;12:297-303.
81. Kanbe N, Tanaka A, Kanbe M, et al. Human mast cells produce matrix metalloproteinase-9. *Eur J Immunol* 1999;29:2645-2649.
82. Lichtinghagen R, Seifert T, Kracke A, et al. Expression of matrix metalloproteinase-9 and its inhibitors in mononuclear blood cells of patients with multiple sclerosis. *J Neuroimmunol* 1999;99:19-26.
83. Clements JM, Cossins JA, Wells GM, et al. Matrix metalloproteinase expression during experimental autoimmune encephalomyelitis and effects of a combined matrix metalloproteinase and tumour necrosis factor-alpha inhibitor. *J Neuroimmunol* 1997;74:85-94.
84. Liedtke W, Cannella B, Mazzaccaro RJ, et al. Effective treatment of models of multiple sclerosis by matrix metalloproteinase inhibitors. *Ann Neurol* 1998;44:35-46.
85. Brenner T, Soffer D, Shalit M, Levi-Schaffer F. Mast cells in experimental allergic encephalomyelitis: characterization, distribution in the CNS and in vitro activation by myelin basic protein and neuropeptides. *J Neurol Sci* 1994;122:210-213.
86. Gallea-Robache S, Morand V, Millet S, et al. A metalloproteinase inhibitor blocks the shedding of soluble cytokine receptors and processing of transmembrane cytokine precursors in human monocytic cells. *Cytokine* 1997;9:340-346.
87. Tsukada N, Miyagi K, Matsuda M, et al. Tumour necrosis factor and interleukin-1 in the CSF and sera of patients with multiple sclerosis. *J Neurol Sci* 1991;104:230-234.
88. Sharief MK, Hentges R. Association between tumour necrosis factor-alpha and disease progression in patients with multiple sclerosis. *N Engl J Med* 1991;325:467-472.
89. Selmaj K, Raine CS, Cross AH. Anti-tumour necrosis factor therapy abrogates autoimmune demyelination. *Ann Neurol* 1991;30:694-700.
90. Cross A, Manning P, Keeling R, et al. Peroxynitrite formation within the central nervous system in active multiple sclerosis. *J Neuroimmunol* 1998;88:45-56.
91. Gottschall PE, Deb S. Regulation of matrix metalloproteinase expressions in astrocytes, microglia and neurons. *Neuroimmunomodulation* 1996;3:69-75.
92. Gulati A, Dhawan KN, Shukla R, et al. Evidence for the involvement of histamine in the regulation of blood-brain barrier permeability. *Pharmacol Res Commun* 1985;17:395-404.
93. Bebo BF Jr, Lee CH, Orr EL, et al. Mast cell-derived histamine and tumour necrosis factor: differences between SJL/J and BALB/c inbred strains of mice. *Immunol Cell Biol* 1996;74:225-230.

94. Yong T, Bebo BF Jr, Sapatino BV, et al. Histamine-induced microvascular leakage in pial venules: differences between the SJL/J and BALB/c inbred strains of mice. *J Neurotrauma* 1994;11:161-171.
95. Weber J, Grise P, Roquebert M, et al. Radioopaque markers transit and anorectal manometry in 16 patients with multiple sclerosis and urinary bladder dysfunction. *Dis Colon Rectum* 1987;30:95-100.
96. Gershon M. *The Second Brain*. New York, NY: Harper Collins;1999:175.
97. Gupta J, Ingegno A, Cook A, Pertschuk L. Multiple sclerosis and malabsorption. *Am J Gastroenterology* 1977;68:560-565.
98. Iarosh OO, Kanevska SA. The characteristics of blood histamine indices and of the pathomorphological changes in gastric mucosa of patients with multiple sclerosis. *Lik Sprava* 1992;1:75-76.
99. Andres MR, Bingham HR. Tubeless gastric analysis with a radio-telemetering pill (Heidelberg capsule). *Can Med Assoc J* 1970;102:1087.
100. Oriente P, Riccio A, Farinaro C, et al. Plasma free aminoacids pattern in progressive systemic sclerosis. *Boll Soc Ital Biol Sper* 1984;60:641-647.
101. Ivanokov AN, Dubrovskaia MK, Karneev AN, Nefedov EP. Spectrum of plasma free amino acids in multiple sclerosis patients. *Zh Nevropatol Psikhiatr Im S S Korsakova* 1984;84:185-189.
102. Qureshi GA, Baig MS. Quantitation of free amino acids in biological samples by high-performance liquid chromatography. Application of the method in evaluating amino acid levels in cerebrospinal fluid and plasma of patients with multiple sclerosis. *J Chromatogr* 1988;459:237-244.
103. Nyhan WL, Hilton S. Histidinuria: defective transport of histidine. *Am J Med Genet* 1992;44:558-561.
104. Palm R, Hallmans G. Zinc and copper in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 1982;45:691-698.
105. Kapaki E, Segditsa J, Papageorgiou C. Zinc, copper and magnesium concentration in serum and CSF of patients with neurological disorders. *Acta Neurol Scand* 1989;79:373-378.
106. Cianciaruso B, Jones MR, Kopple JD. Histidine, an essential amino acid for adult dogs. *J Nutr* 1981;111:1074-1084.
107. Schuschke DA, Saari JT, Miller FN. The role of the mast cell in acute inflammatory responses of copper-deficient rats. *Agents Actions* 1994;42:19-24.
108. Sharma SC, Jande MB. Inhibition of mast cell histamine release by copper. *Arch Int Pharmacodyn Ther* 1989;299:254-268.
109. Jackson MC, Lenney JF. The distribution of carnosine and related dipeptides in rat and human tissues. *Inflamm Res* 1996;45:132-135.
110. Rajendran V, Berteloot A, Ishikawa Y, et al. Transport of carnosine by mouse intestinal brush-border membrane vesicles. *Biochim Biophys Acta* 1984;778:443-448.
111. Arnould JM. In vitro demonstration of histamine biosynthesis from carnosine by kidneys of pregnant mice. *Can J Physiol Pharmacol* 1987;65:70-74.
112. Fitzpatrick JC, Fisher H, Flancbaum L. Mobilization of renal carnosine and histidine to histamine during compound-48/80-induced shock. *Nephron* 1991;59:299-303.
113. Flancbaum L, Fitzpatrick JC, Brotman DN, et al. The presence and significance of carnosine in histamine-containing tissues of several mammalian species. *Agents Actions* 1990;31:190-196.
114. Babizhayev MA, Seguin MC, Gueyne J, et al. L-carnosine (beta-alanyl-L-histidine) and carcinine (beta-alanylhistamine) act as natural antioxidants with hydroxyl-radical-scavenging and lipid-peroxidase activities. *Biochem J* 1994;304:509-516.
115. Boldyrev A, Abe H. Metabolic transformation of neuropeptide carnosine modifies its biological activity. *Cell Mol Neurobiol* 1999 Feb;19:163-175.
116. MacFarlane N, McMurray J, O'Dowd JJ, et al. Synergism of histidyl dipeptides as antioxidants. *J Mol Cell Cardiol* 1991;23:1205-1207.
117. Wassif WS, Sherwood RA, Amir A, et al. Serum carnosinase activities in central nervous system disorders. *Clin Chim Acta* 1994;225:57-64.
118. Bando K, Ichihara K, Shimotsuji T, et al. Reduced serum carnosinase activity in hypothyroidism. *Ann Clin Biochem* 1986;23:190-194.
119. Kiessling WR, Pflughaupt KW, Haubitz I, et al. Thyroid function in multiple sclerosis. *Acta Neurol Scand* 1980;62:255-258.

120. Voskoboyev AI, Artsukevich IM, Ostrovsky YM. The functional role of histidine and sulfhydryl groups in rat liver thiamine pyrophosphokinase. *Acta Vitaminol Enzymol* 1983;5:105-113.
121. Rapala-Kozik M, Kozik A. Mechanism of ligand-protein interaction in plant seed thiamine-binding proteins. Preliminary chemical identification of amino acid residues essential for thiamine binding to the buckwheat seed protein. *Biochimie* 1996;78:77-84.
122. Harris RA, Hawes JW, Popov KM, et al. Studies on the regulation of the mitochondrial alpha-ketoacid dehydrogenase complexes and their kinases. *Adv Enzyme Regul* 1997;37:271-293.
123. Khramtsov AV, Morozov IA, Martinchik AN. Ultrastructure of the rat gastric mucosal parietal cells in thiamine deficiency. *Vopr Pitan* 1979;4:48-52.
124. DeLack E, personal communication.
125. Kamalian N, Keesey RE, Zu R, et al. Lateral hypothalamic demyelination and cachexia in a case of "malignant" multiple sclerosis. *Neurology* 1975;25:25-30.
126. Salido G, Lennard R, Singh J, Pariente J. Histamine-evoked amylase secretion is associated with small changes in calcium mobilization in isolated guinea-pig pancreas. *Exp Physiol* 1990;75:263-266.
127. Nguyen TD, Okolo CN, Moody MW. Histamine stimulates ion transport by dog pancreatic duct epithelial cells through H1 receptors. *Am J Physiol* 1998;275:G76-G84.
128. Singh J, Pariente JA, Salido GM. The physiological role of histamine in the exocrine pancreas. *Inflamm Res* 1997;46:159-165.
129. Panula P, Kaartinen M, Macklin M, Costa E. Histamine-containing peripheral neuronal and endocrine systems. *J Histochem Cytochem* 1985;33:933-941.
130. Singaram C, Ashraf W, Gaumnitz EA, et al. Dopaminergic defect of enteric nervous system in Parkinson's disease patients with chronic constipation. *Lancet* 1995;346:861-864.
131. Wakabayashi K, Takahashi H. Neuropathology of autonomic nervous system in Parkinson's disease. *Eur Neurol* 1997;38:2-7.
132. Puurunen J. Centrally applied histamine increases gastric acid secretion in rats. *Naunyn Schmiedebergs Arch Pharmacol* 1988;338:96-98.
133. Wang ZL, Lu GQ. Effect of intraventricular administration of histamine and its receptor agonists on pentagastrin-induced gastric acid secretion in rats. *Sheng Li Hsueh Pao* 1992;44:261-268.
134. Barocelli E, Ballabeni V, Chiavarini M, Impicciatore M. R-alpha-methylhistamine-induced inhibition of gastric acid secretion in pylorus-ligated rats via central histamine H3 receptors. *Br J Pharmacol* 1995;115:1326-1330.
135. Tache Y, Goto Y, Hamel D, et al. Mechanisms underlying intracisternal TRH-induced stimulation of gastric acid secretion in rats. *Regul Pept* 1985;13:21-30.
136. Okumura T, Uehara A, Okumura K, et al. Inhibition of gastric pepsin secretion by peripherally or centrally injected interleukin-1 in rats. *Biochem Biophys Res Commun* 1990;167:956-961.
137. Gerber DA, Tanenbaum L, Ahrens M. Free serum histidine levels in patients with rheumatoid arthritis and control subjects following an oral load of free L-histidine. *Metabolism* 1976;25:655-657.
138. Said HM, Mohammadkhani R. Involvement of histidine residues and sulfhydryl groups in the function of the biotin transport carrier of rabbit intestinal brush-border membrane. *Biochim Biophys Acta* 1992;1107:238-244.
139. Said HM, Mohammadkhani R. Folate transport in intestinal brush border membrane: involvement of essential histidine residue(s). *Biochem J* 1993;290:237-240.
140. Suojaranta-Ylinen R, Hendolin H, Tuomisto L. The effects of morphine, morphine plus scopolamine, midazolam and promethazine on cerebrospinal fluid histamine concentration and postoperative analgesic consumption. *Agents Actions* 1991;33:212-214.
141. Kiviranta T, Tuomisto L, Airaksinen EM. Histamine in cerebrospinal fluid of children with febrile convulsions. *Epilepsia* 1995;36:276-280.
142. Rozniecki JJ, Hauser SL, Stein M, et al. Elevated mast cell tryptase in cerebrospinal fluid of multiple sclerosis patients. *Ann Neurol* 1995;37:63-66.
143. Tuomisto L, Kilpelainen H, Riekkinen P. Histamine and histamine-N-methyltransferase in the CSF of patients with multiple sclerosis. *Agents Actions* 1983;13:255-257.

144. Molnar G, Moldavan J. Histamine content of the cerebrospinal fluid in multiple sclerosis: A preliminary communication. *Acta Med Acad Sci Hung* 1966;22:271-274.
145. Rai V, Dubey S, Singh R, Udupa K. Histamine and histaminase levels in blood. *Indian J Med Res* 1977;66:150-154.
146. Mammen GJ, Pennefather JN. The actions of histamine on the separated layers of the guinea-pig uterus: the influence of ovarian steroids. *Agents Actions* 1988;24:49-55.
147. Bodis J, Tinneberg HR, Schwarz H, et al. The effect of histamine on progesterone and estradiol secretion of human granulosa cells in serum-free culture. *Gynecol Endocrinol* 1993;7:235-239.
148. Krishna A, Terranova PF. Compartmentalized mast cell degranulations in the ovarian hilum, fat pad, bursa and blood vessel regions of the cyclic hamster: relationships to ovarian histamine and blood flow. *Acta Anat (Basel)* 1991;141:18-25.
149. Fisk J, Pontefract A, Ritvo P. The impact of fatigue on patients with multiple sclerosis. *Canada J Neur Sci* 1994;21:9-14.
150. Brenneis M, Harrer G, Selzer H. On the temperature sensitivity of multiple sclerosis patients. *Fortschr Neurol Psychiatr Genzgeb* 1979;47:320-325.
151. Guthrie TC, Nelson DA. Influence of temperature changes on multiple sclerosis: critical review of mechanisms and research potential. *J Neurol Sci* 1995;129:1-8.
152. Knigge U, Alsbjorn B, Thuesen B, et al. Temporal responses of cutaneous blood flow and plasma catecholamine concentrations to histamine H1- or H2-receptor stimulation in man. *Eur J Clin Pharmacol* 1988;33:613-617.
153. Good P. Histamine vasodilation — a forgotten effective treatment for multiple sclerosis. Unpublished manuscript. Author contact: Teslaphile2000@aol.com.
154. McCormick DA. Functional properties of a slowly inactivating potassium current in guinea pig dorsal lateral geniculate relay neurons. *J Neurophysiol* 1991;66:1176-1189.
155. Bostock H, Sears TA, Sherratt RM. The effects of 4-aminopyridine and tetraethylammonium ions on normal and demyelinated nerve fibers. *J Physiol* 1981;313:301-315.
156. Targ EF, Kocsis JD. 4-aminopyridine leads to restoration of conduction in demyelinated rat sciatic nerve. *Brain Res* 1985;328:358-361.
157. Davis F, Stefoski D, Rush J. Orally administered 4-Aminopyridine improves clinical signs in multiple sclerosis. *Ann Neurol* 1990;27:186-192.
158. Bever CT, Young D, Anderson PA, et al. The effects of 4-aminopyridine in multiple sclerosis patients: results of a randomized, placebo-controlled, double-blind, concentration-controlled, crossover trial. *Neurology* 1994;44:1054-1059.
159. Reiner PB, Kamondi A. Mechanisms of antihistamine-induced sedation in the human brain: H1 receptor activation reduces a background leakage potassium current. *Neuroscience* 1994;59:579-588.
160. Gorelova N, Reiner PB. Histamine depolarizes cholinergic septal neurons. *J Neurophysiol* 1996;75:707-714.
161. Li Z, Hatton GI. Histamine-induced prolonged depolarization in rat supraoptic neurons: G-protein-mediated, Ca(2+)-independent suppression of K⁺ leakage conductance. *Neuroscience* 1996;70:145-158.
162. Jafri MS, Moore KA, Taylor GE, Weinreich D. Histamine H1 receptor activation blocks two classes of potassium current, I_{K(rest)} and IAHP, to excite ferret vagal afferents. *J Physiol* 1997;503:533-546.
163. Cordoba-Rodriguez R, Moore KA, Kao JP, Weinreich D. Calcium regulation of a slow post-spike hyperpolarization in vagal afferent neurons. *Proc Natl Acad Sci USA* 1999;96:7650-7657.
164. Glover WE. Potentiation of vasoconstrictor responses by 3- and 4-aminopyridine. *Br J Pharmacol* 1978;63:577-585.
165. Rodriguez FJ, Lluch M, Dot J, et al. Histamine modulation of glutamate release from hippocampal synaptosomes. *Eur J Pharmacol* 1997;323:283-286.
166. Bourne HR, Melmon KI, Lichtenstein LM. Histamine augments leukocyte adenosine 3',5'-monophosphate and blocks antigenic histamine release. *Science* 1971;173:743-745.
167. Banu Y, Watanabe T. Augmentation of antigen receptor-mediated responses by histamine H1 receptor signaling. *J Exp Med* 1999;189:673-682.
168. Rezai AR, Salazar-Gonzalez JF, Martinez-Maza O, et al. Histamine blocks interleukin 2 (IL-2) gene expression and regulates IL-2 receptor expression. *Immunopharmacol Immunotoxicol* 1990;12:345-362.

169. Hellstrand K, Hermodsson S, Naredi P, et al. Histamine and cytokine therapy. *Acta Oncol* 1998;37:347-353.
170. Elenkov IJ, Webster E, Papanicolaou DA, et al. Histamine potently suppresses human IL-12 and stimulates IL-10 production via H2 receptors. *J Immunol* 1998;161:2586-2593.
171. van der Pouw Kraan TC, Snijders A, Boeije LC, et al. Histamine inhibits the production of interleukin-12 through interaction with H2 receptors. *J Clin Invest* 1998;102:1866-1873.
172. Lichtenstein L, Gillespie E. Inhibition of histamine release by histamine controlled by H2 receptor. *Nature* 1973;244:297-298.
173. Sullivan TJ, Parker KL, Eisen SA, et al. Modulation of cyclic AMP in purified rat mast cells. II. Studies on the relationship between intracellular cyclic AMP concentrations and histamine release. *J Immunol* 1975;114:1480-1485.
174. Kurosawa M, Kobayashi S. Changes in protein kinase C activity during histamine release from activated rat mast cells. *Allergy* 1989;44:226-232.
175. Teraoka H, Akiba H, Takai R, et al. Inhibitory effects of caffeine on Ca²⁺ influx and histamine secretion independent of cAMP in rat peritoneal mast cells. *Gen Pharmacol* 1997;28:237-243.
176. Dinter H, Onuffer J, Faulds D, et al. Phosphodiesterase type IV inhibitors in the treatment of MS. *J Mol Med* 1997;75:95-102.
177. Sommer N, Martin R, McFarland HF, et al. Therapeutic potential of phosphodiesterase type 4 inhibition in chronic autoimmune demyelinating disease. *J Neuroimmunol* 1997;79:54-61.
178. Bielekova B, Lincoln A, McFarland H, Martin R. Therapeutic potential of phosphodiesterase-4 and β 3 inhibitors in Th1-mediated autoimmune diseases. *J Immunol* 2000;164:1117-1124.
179. Rieckmann P, Weber F, Gunther A, et al. Pentoxifylline, a phosphodiesterase inhibitor, induces immune deviation in patients with multiple sclerosis. *J Neuroimmunol* 1996;64:193-200.
180. Ulloa ER, Zaninovich AA. Effects of histamine H1- and H2-receptor antagonists on thyrotrophin secretion in the rat. *J Endocrinol* 1986;111:175-180.
181. Onaya T, Hashizume K, Sato A, et al. Evidence for the existence of a histamine H2-receptor in the mouse thyroid. *Endocrinology* 1977;100:61-66.
182. Swank RL, Roth JG, Woody DC Jr. Cerebral blood flow and red cell delivery in normal subjects and in multiple sclerosis. *Neurol Res* 1983;5:37-59.
183. Brooks DJ, Leenders KL, Head G, et al. Studies on regional cerebral oxygen utilisation and cognitive function in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 1984;47:1182-1191.
184. Sun X, Tanaka M, Kondo S. Clinical significance of reduced cerebral metabolism in multiple sclerosis: a combined PET and MRI study. *Ann Nucl Med* 1998;12:89-94.
185. Lycke J, Wikkelso C, Bergh AC, et al. Regional cerebral blood flow in multiple sclerosis measured by single photon emission tomography with technetium-99m hexamethylpropyleneamine oxime. *Eur Neurol* 1993;33:163-167.
186. Sercombe R, Verrecchia C, Philipson V, et al. Histamine-induced constriction and dilation of rabbit middle cerebral arteries in vitro: role of the endothelium. *Blood Vessels* 1986;23:137-153.
187. Rosenblum WI, Nelson GH, Weinbrecht P. Histamine elicits competing endothelium-dependent constriction and endothelium-independent dilation in vivo in mouse cerebral arterioles. *Stroke* 1990;21:305-309.
188. Toda N. Mechanism underlying responses to histamine of isolated monkey and human cerebral arteries. *Am J Physiol* 1990;258:H311-H317.
189. Horton, BT. The clinical use of histamine. *Postgrad Med* 1951;9:1-23.
190. Gonen O, Catalaa I, Babb JS, et al. Total brain N-acetylaspartate: a new measure of disease load in MS. *Neurology* 2000;54:15-19.
191. Burlina AP, Ferrari V, Facci L, et al. Mast cells contain large quantities of secretagogue-sensitive N-acetylaspartate. *J Neurochem* 1997;69:1314-1317.